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## New data and update in the *Panaeolus papilionaceus* complex, first record of *P. punjabensis* and new finds of *P. sylvaticus* in the Dominican Republic

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### Key words:

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Agaricales  
Galeropsidaceae  
Caribbean  
taxonomy  
phylogeny

**Abstract:** *Panaeolus punjabensis*, recently described from Pakistan, is first documented from the Dominican Republic based on morphological description and molecular analyses (ITS sequence and phylogram). An updating on the *Panaeolus papilionaceus* complex is reported based on new molecular evidence on *P. papilionaceus* var. *parvisporus*. New finds of *P. sylvaticus* in the Dominican Republic are documented with photographs and morphological notes.

## INTRODUCTION

In the family Galeropsidaceae, the genus *Panaeolus* has a cosmopolitan diffusion and is mainly composed of fimicolous to nitrophilous species (Voto & Angelini 2024). It is widely spread in the Dominican Republic where the habit of raising numerous herbivorous farm animals (such as cows, sheep, goats, horses and donkeys) exclusively on pasture in grassy unwooded spaces creates a particularly favourable habitat.

To the four Dominican species of *Panaeolus* (*P. antillarum*, *P. mexicanus*, *P. pantropicalis* sp. nov., *P. sylvaticus*) documented in previous works (Voto & Angelini 2021; Angelini & Voto 2023; Voto & Angelini 2024) we add now *P. punjabensis*.

In the second part of the work, following very recent new molecular evidence, we propose an update of our previous work regarding nomenclatural issues in the *Panaeolus papilionaceus* complex.

Finally, new photographs of *P. sylvaticus* are published showing more chromatic aspects of its pileus.

## MATERIALS AND METHODS

### Morphology

The collections of *Panaeolus punjabensis* and *P. sylvaticus* were photographed when fresh *in situ* by C. Angelini using a digital camera Nikon Coolpix 8400 and subsequently dried; macroscopic characters were observed by C. Angelini on fresh material; microscopic characters were studied, described and photographed by P. Voto on dried material revived with 10% NH<sub>4</sub>OH or 5% KOH and in some cases using Congo red as mounting medium for imaging. All collections are housed in C. Angelini's personal herbarium and will successively be transferred to the herbarium JBSD (Jardín Botánico Nacional Dr. Rafael M. Moscoso, Santo Domingo, Dominican Republic). The authors of the images in the figures are shown in square brackets.

### DNA extraction, PCR and DNA sequencing

Total DNA was extracted from dry specimens employing a modified protocol based on Murray & Thompson (1980). PCR reactions (Mullis & Faloona 1987) included 35 cycles with an annealing temperature of 54 °C. The primers ITS1F and ITS4 (White *et al.* 1990; Gardes & Bruns 1993) were employed to amplify the ITS rDNA region. PCR products were checked in 1% agarose gels, and amplicons were sequenced with one or both PCR primers. Sequences were corrected to remove reading errors in chromatograms. An alignment of ITS rDNA

sequences related to the one obtained from the sample studied was assembled, using the dataset employed before (Voto & Angelini 2024) as well as selected sequences produced by Consiglio & Marchetti (2023).

### Sequence alignment and phylogenetic analyses

Sequences first were aligned in MEGA 5.0 (Tamura *et al.* 2011) with its Clustal W application and then realigned manually as needed to establish positional homology. Aligned sequences were loaded in MrBayes 3.2.6 (Ronquist *et al.* 2012) and subjected to Bayesian analysis (one partition, GTR+G+I model, two simultaneous runs, four chains, temperature set to 0.2, sampling every 100th generation) until the average split frequencies between the simultaneous runs fell below 0.01 after 14.55 M generations. Finally, a full search for the best-scoring maximum likelihood tree was performed in RAXML 8.2.12 (Stamatakis 2014) using the standard search algorithm (same partitions, GTRGAMMAI model, 2000 bootstrap replications). The significance threshold was set above 0.95 for posterior probability (PP) and 70% bootstrap proportions (BP).

## RESULTS

### Phylogenetic analyses

Our phylogram demonstrates that *Panaeolus punjabensis* is a cosmopolitan species that, besides Asia (Pakistan) and Central America (Dominican Republic) is also present in Africa (Egypt and Namibia).

Concerning the *Panaeolus papilionaceus* complex, the results obtained in our previous work (Voto & Angelini 2024) are confirmed with the corrections, thanks to the inclusion of the holotype of *Panaeolus papilionaceus* var. *parvisporus*, that the clade there identified as *P. parvisporus* must be named *P. retirugis* and that this name must be treated as autonomous, not as a synonym of *P. papilionaceus*.

**Table 1.** Data of specimens used in this study. The column 'Current name' is used for the current name of species which are synonymised (in bold) or vouchers which are misidentified (not in bold). Current names proposed here or in Voto & Angelini (2024) are in red, those proposed by other authors are identified by asterisks (\*: Malysheva *et al.* 2019, \*\*: Asif *et al.* 2023). The accession numbers of the newly generated sequences are in bold blue. Bold capital letters after species names represent Holotype (T), Epitype (E), Paratypes (P) and Lectotypes (L)

species name in GenBank	current name	voucher/origin	ITS
<i>Galeropsis bispora</i> T	<i>P. plantaginiformis</i> *	LE 2863/Uzbekistan	MK397580
<i>G. desertorum</i>	<i>P. desertorum</i> *	NL-1863/Hungary	JX968154
<i>Gastrocybe iberica</i> T	<i>P. desertorum</i> *	AH 9990/Spain	MK397542
<i>G. iberica</i> P	<i>P. desertorum</i> *	AH 9993/Spain	MK397543
<i>G. iberica</i>	<i>P. desertorum</i> *	AH 10396/Spain	MK397544
<i>G. iberica</i>	<i>P. desertorum</i> *	AH 42860/Spain	MK397545
<i>G. iberica</i>	<i>P. desertorum</i> *	AH 10493/Spain	MK397546
<i>Panaeolus acuminatus</i> E		Gerhardt 83049 (B)/Germany	PP447476
<i>P. acuminatus</i>		CBS:270.47/?	MH856251
<i>P. acuminatus</i>		AMB 20065/Italy	PP447475
<i>P. acuminatus</i>		inaturalist.org/observations/141352004/USA	OQ372246
<i>P. acuminatus</i>		SGL09/China	OR035540
<i>P. acuminatus</i>		CBS 269.47/?	MH856250
<i>P. acuminatus</i>	<i>P. olivaceus</i>	PRM 935914/Czech Republic	MW352021
<i>P. acuminatus</i>	<i>P. fimicola</i>	4084/Italy	JF908518
<i>P. alcis</i>		Mushroom Observer #88085/Sweden	KM982723
<i>P. alcis</i>		SAT-14-239-20/USA(Alaska)	MW597122
<i>P. antillarum</i>		SFSU:DED7874/Thailand	MF497585
<i>P. antillarum</i>		PAD H0061942/Dominican Republic	<b>PP590043</b>

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<i>P. antillarum</i>		CORT:013830/Dominican Republic	MF497586
<i>P. antillarum</i>		JL27/China	ON059337
<i>P. antillarum</i>		EGDA-N15/Egypt	ON024905
<i>P. antillarum</i>		BR5020167127933/Togo	OR035499
<i>P. antillarum</i>		WANG 140007/Taiwan	KR998382
<i>P. antillarum</i>		FLAS-F-69480/USA	OQ746434
<i>P. antillarum</i>		DQS36F/Philippines	MZ735416
<i>P. antillarum</i>		HYW22/Thailand	OR035522
<i>P. antillarum</i>		HYW21/Thailand	OR035520
<i>P. antillarum</i>	<i>P. semiovatus</i>	748/Italy	JF908515
<i>P. axfordii</i> T		MFLU:19-2367/China	NR_169700
<i>P. bisporus</i>	<i>P. cyanescens</i>	KaiR95/Benin	MT110229
<i>P. bisporus</i>	<i>P. cyanescens</i>	MushroomObserver.org/188954/USA	MG966283
<i>P. bisporus</i>	<i>P. cyanescens</i>	HYW197/China	OR035518
<i>P. bisporus</i>	<i>P. cyanescens</i>	T2507/China	OR035542
<i>P. cambodginiens</i>	<i>P. cyanescens</i>	NBRC-30222/Japan?	AB158633
<i>P. campanulatus</i>	<i>P. papilionaceus</i>	10141/Italy	JF908522
<i>P. campanulatus</i>	<i>P. papilionaceus</i>	Mushroom2/China	MT451920
<i>P. campanulatus</i>	<i>P. cinctulus</i>	/China	JF961376
<i>P. castaneifolius</i>	<i>P. foenisecii</i>	Mushroom Observer 90428/USA	KX010428
<i>P. cinctulus</i> E		Gerhardt 83052/Germany	PP447483
<i>P. cinctulus</i>		iNAT:56796374/USA	OQ147191
<i>P. cinctulus</i>		iNAT:126276059/USA	OP751540
<i>P. cinctulus</i>		S.D. Russell ONT iNaturalist 130442846/USA	OP549138
<i>P. cinctulus</i>		PRM 935916, Mushroom Observer 204889/Italy	MW352022
<i>P. cinctulus</i>		Mushroom Observer # 321948/USA	MH590045
<i>P. cinctulus</i>		OMDL K. Canan iNaturalist # 169757989/USA	OR987159
<i>P. cinctulus</i>		OMDL K. Canan iNaturalist # 170057671/USA	OR987260
<i>P. cinctulus</i>		MCVE 1084/Italy	PP447482
<i>P. cinctulus</i>		iNat86856915/USA	OQ389417
<i>P. cinctulus</i>		BR5020180462561/Netherlands	OR035501
<i>P. cinctulus</i>		HYW186/China	OR035517
<i>P. cyanescens</i>		AF163/China	OQ450458
<i>P. cyanescens</i>		D36/China	KT002152
<i>P. cyanescens</i>		MHHNU 31855/China	OP862802
<i>P. cyanescens</i>		SIV1/India	MK855517
<i>P. cyanescens</i>		FS1/India	MK855516
<i>P. cyanescens</i>		SIV2/India	MK855518
<i>P. cyanescens</i>		LAH37983_Panaeolus_cyanescens_sm38/PAK	OR668690
<i>P. cyanescens</i>		LAH37985_Panaeolus_cyanescens_sm40/PAK	OR668692
<i>P. cyanescens</i>		MES-4248/USA	OR664094
<i>P. cyanescens</i>		BP17M/Philippines	OR062405
<i>P. cyanescens</i>		1S/Croatia?	KU640168
<i>P. cyanescens</i>		18S/?	HM035084

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<i>P. cyanescens</i>		0709305/JG/France	PP447485
<i>P. cyanescens</i>		AMB 20070/Italy	PP447484
<i>P. cyanescens</i> var.	<i>P. cyanescens</i>	n. 6576 AQU/Italy	EU834287
<i>P. desertorum</i>		LE 313090/Russia	MK397566
<i>P. desertorum</i>		LE 313250/Russia	MK397568
<i>P. desertorum</i>		LE 313091/Greece	MK397567
<i>P. desertorum</i>		B. Dima BG-2022-10-19-1/Hungary	PP447487
<i>P. desertorum</i>		GB-0073426/Hungary	PP447486
<i>P. desertorum</i>	<i>P. plantaginiformis</i>	LE 2864/Uzbekistan	MH055384
<i>P. desertorum</i>	<i>P. plantaginiformis</i>	LE 2865/Uzbekistan	MH055383
<i>P. dunensis</i>		AMB 20210/France	PP447489
<i>P. dunensis</i>		AMB 20211/Italy	PP447490
<i>P. fimicola</i> E		Gerhardt 75349 (B)/Germany	PP447491
<i>P. fimicola</i> E		Gerhardt 75349 (B)/Germany	PP447492
<i>P. fimicola</i>	<i>P. semiovatus</i>	NSK 1017274/Russia	OR242695
<i>P. fimicola</i>	<i>P. semiovatus</i>	Mushroom6/China	MT451924
<i>P. fimicola</i>	<i>P. semiovatus</i>	20180624002/China	MT347601
<i>P. fimicola</i>	<i>P. semiovatus</i>	474/Italy	JF908514
<i>P. fimicola</i>	<i>P. semiovatus</i>	HMJAU66147/China	OQ927089
<i>P. foeniseeii</i>		T-790/USA	KC176293
<i>P. foeniseeii</i>		S.D. Russell MycoMap # 5430/USA	ON561649
<i>P. foeniseeii</i>		S.D. Russell ONT iNaturalist # 141290781/USA	OQ297023
<i>P. foeniseeii</i>		iNAT:21657597/USA	OM212934
<i>P. foeniseeii</i>		S.D. Russell MycoMap # 5431/USA	ON561650
<i>P. foeniseeii</i>		OMDL K. Canan iNaturalist # 172543719/USA	OR732083
<i>P. foeniseeii</i>		OMDL K. Canan iNaturalist # 169180865/USA	OR825597
<i>P. foeniseeii</i>		OMDL K. Canan iNaturalist # 171232504/USA	OR785929
<i>P. foeniseeii</i>		S.D. Russell ONT iNaturalist 118906067/USA	OP749347
<i>P. foeniseeii</i>		S.D. Russell ONT iNaturalist # 121782791/USA	OP470404
<i>P. foeniseeii</i>		OMDL K. Canan iNaturalist # 170924896/USA	OR732074
<i>P. foeniseeii</i>		JLF9301 iNaturalist # 81665784/USA	OQ859921
<i>P. foeniseeii</i>		S.D. Russell MycoMap # 5536/USA	ON561653
<i>P. foeniseeii</i>		S.D. Russell ONT iNaturalist 127360021/USA	OP549249
<i>P. foeniseeii</i>		GS6/China	OR035507
<i>P. foeniseeii</i>		M8/Hungary	OQ029266
<i>P. foeniseeii</i>		AMB 20071/Italy	PP447493
<i>P. foeniseeii</i>		AMB 20072/Italy	PP447494
<i>P. foeniseeii</i>		OTA:71571/New Zealand	OQ064958
<i>P. foeniseeii</i>		6643/Italy	JF908520
<i>P. foeniseeii</i>		K(M):250281/United Kingdom	MZ159698
<i>P. foeniseeii</i>		BR5020160357160/Belgium	OR035496
<i>P. foeniseeii</i>		CBS 142.40/?	MH856067
<i>P. foeniseeii</i>		CBS 143.40/?	MH856068

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<i>P. foeniseccii</i>	<i>P. sp.</i>	CBS 251.37/?	MH855904
<i>P. foeniseccii</i>	<i>P. antillarum</i>	NSK 1017353/Russia	OR364524
<i>P. foeniseccii</i>	<i>P. antillarum</i>	UOC-KAUNP-MK62/Sri Lanka	KP764810
<i>P. foeniseccii</i>	<i>P. antillarum</i>	UOC KAUNP K01/Sri Lanka	KR867660
<i>P. foeniseccii</i>	<i>P. antillarum</i>	PC14/Philippines	OK446756
<i>P. fraxinophilus</i>	<i>P. dunensis</i>	OMDL K. Canan iNaturalist # 170758482/USA	OR987324
<i>P. fraxinophilus</i>	<i>P. dunensis</i>	MushroomObserver.org/455364/USA	OL629088
<i>P. desertorum</i>		LE 313090/Russia	MK397566
<i>P. desertorum</i>		LE 313250/Russia	MK397568
<i>P. desertorum</i>		LE 313091/Greece	MK397567
<i>P. desertorum</i>		B. Dima BG-2022-10-19-1/Hungary	PP447487
<i>P. desertorum</i>		GB-0073426/Hungary	PP447486
<i>P. desertorum</i>	<i>P. plantaginiformis</i>	LE 2864/Uzbekistan	MH055384
<i>P. desertorum</i>	<i>P. plantaginiformis</i>	LE 2865/Uzbekistan	MH055383
<i>P. dunensis</i>		AMB 20210/France	PP447489
<i>P. dunensis</i>		AMB 20211/Italy	PP447490
<i>P. fimicola</i> E		Gerhardt 75349 (B)/Germany	PP447491
<i>P. fimicola</i> E		Gerhardt 75349 (B)/Germany	PP447492
<i>P. fimicola</i>	<i>P. semiovatus</i>	NSK 1017274/Russia	OR242695
<i>P. fimicola</i>	<i>P. semiovatus</i>	Mushroom6/China	MT451924
<i>P. fimicola</i>	<i>P. semiovatus</i>	20180624002/China	MT347601
<i>P. fimicola</i>	<i>P. semiovatus</i>	474/Italy	JF908514
<i>P. fimicola</i>	<i>P. semiovatus</i>	HMJAU66147/China	OQ927089
<i>P. foeniseccii</i>		T-790/USA	KC176293
<i>P. foeniseccii</i>		S.D. Russell MycoMap # 5430/USA	ON561649
<i>P. foeniseccii</i>		S.D. Russell ONT iNaturalist # 141290781/USA	OQ297023
<i>P. foeniseccii</i>		iNAT:21657597/USA	OM212934
<i>P. foeniseccii</i>		S.D. Russell MycoMap # 5431/USA	ON561650
<i>P. foeniseccii</i>		OMDL K. Canan iNaturalist # 172543719/USA	OR732083
<i>P. foeniseccii</i>		OMDL K. Canan iNaturalist # 169180865/USA	OR825597
<i>P. foeniseccii</i>		OMDL K. Canan iNaturalist # 171232504/USA	OR785929
<i>P. foeniseccii</i>		S.D. Russell ONT iNaturalist 118906067/USA	OP749347
<i>P. foeniseccii</i>		S.D. Russell ONT iNaturalist # 121782791/USA	OP470404
<i>P. foeniseccii</i>		OMDL K. Canan iNaturalist # 170924896/USA	OR732074
<i>P. foeniseccii</i>		JLF9301 iNaturalist # 81665784/USA	OQ859921
<i>P. foeniseccii</i>		S.D. Russell MycoMap # 5536/USA	ON561653
<i>P. foeniseccii</i>		S.D. Russell ONT iNaturalist 127360021/USA	OP549249
<i>P. foeniseccii</i>		GS6/China	OR035507
<i>P. foeniseccii</i>		M8/Hungary	OQ029266
<i>P. foeniseccii</i>		AMB 20071/Italy	PP447493
<i>P. foeniseccii</i>		AMB 20072/Italy	PP447494
<i>P. foeniseccii</i>		OTA:71571/New Zealand	OQ064958
<i>P. foeniseccii</i>		6643/Italy	JF908520

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<i>P. foeniseccii</i>		BR5020160357160/Belgium	OR035496
<i>P. foeniseccii</i>		CBS 142.40/?	MH856067
<i>P. foeniseccii</i>		CBS 143.40/?	MH856068
<i>P. foeniseccii</i>	<i>P. sp.</i>	CBS 251.37/?	MH855904
<i>P. foeniseccii</i>	<i>P. antillarum</i>	NSK 1017353/Russia	OR364524
<i>P. foeniseccii</i>	<i>P. antillarum</i>	UOC-KAUNP-MK62/Sri Lanka	KP764810
<i>P. foeniseccii</i>	<i>P. antillarum</i>	UOC KAUNP K01/Sri Lanka	KR867660
<i>P. foeniseccii</i>	<i>P. antillarum</i>	PC14/Philippines	OK446756
<i>P. fraxinophilus</i>	<i>P. dunensis</i>	OMDL K. Canan iNaturalist # 170758482/USA	OR987324
<i>P. fraxinophilus</i>	<i>P. dunensis</i>	MushroomObserver.org/455364/USA	OL629088
<i>P. guttulatus</i>		STA5/Iraq	LC458688
<i>P. guttulatus</i>		AMB n. 18101/Italy	KU725993
<i>P. guttulatus</i>		AMB 20073/Italy	PP447495
<i>P. guttulatus</i> var.		AMB 18102/Italy	KU725994
<i>P. mediterraneus</i> T		AMB 20075/Italy	PP447497
<i>P. mediterraneus</i> P		AMB 20074/Italy	PP447496
<i>P. mexicanus</i>		ANGE1557/Dominican Republic	MZ856314
<i>P. cf. olivaceus</i>	<i>P. olivaceus</i>	MushroomObserver.org/158389/USA	MF629829
<i>P. olivaceus</i>		inaturalist.org/observations/141678308/USA	OQ318240
<i>P. olivaceus</i>		Pan.Olivaceus-ITS1/USA	OQ318238
<i>P. olivaceus</i>		MushroomObserver.org/89608/USA	MH285992
<i>P. olivaceus</i>	<i>P. dunensis</i>	UBC F-32268/Canada	MF955153
<i>P. olivaceus</i>	<i>P. sp.</i>	AMB 20076/Italy	PP447498
<i>P. olivaceus</i>	<i>P. sp.</i>	139/Iran	MH593015
<i>P. pantropicalis</i> T		JBSD 130972/Rep. Dominicana	<a href="#">PP590037</a>
<i>P. pantropicalis</i> P		PAD H0061940/Rep. Dominicana	<a href="#">PP590036</a>
<i>P. pantropicalis</i> P		PAD H0061941/Rep. Dominicana	<a href="#">PP590038</a>
<i>P. pantropicalis</i> P		PERTH 09605894/Australia	<a href="#">PP590039</a>
<i>P. papilionaceus</i> E		Gerhardt 87085 (B)/Germany	PP447500
<i>P. papilionaceus</i> E		Gerhardt 87085 (B)/Germany	PP447499
<i>P. papilionaceus</i>		iNAT:22477730/USA	OM338968
<i>P. papilionaceus</i>		MushroomObserver.org/312080/USA	MH100727
<i>P. papilionaceus</i>		MushroomObserver.org/312173/USA	MH100681
<i>P. papilionaceus</i>		Mushroom Observer 428579/USA	MW633031
<i>P. papilionaceus</i>		S.D. Russell MycoMap # 5533/USA	ON245337
<i>P. papilionaceus</i>		Montri-76/Switzerland?	MK028487
<i>P. papilionaceus</i>		Mushroom Observer # 114447/Mexico	MF628989
<i>P. papilionaceus</i>		STA2/Iran	LC458685
<i>P. papilionaceus</i>		RA400/Iraq	MH632116
<i>P. papilionaceus</i>	<i>P. detriticola</i> T	PERTH 08944954/Australia	MT571659
<i>P. papilionaceus</i>	<i>P. sp.</i>	MHHNU31392/China	MK439503
<i>P. papilionaceus</i>	<i>P. sp.</i>	HNL501769/Lao	UDB033926

**New data and update in the *Panaeolus papilionaceus* complex, first record of *P. punjabensis* and new finds of *P. sylvaticus* in the Dominican Republic**

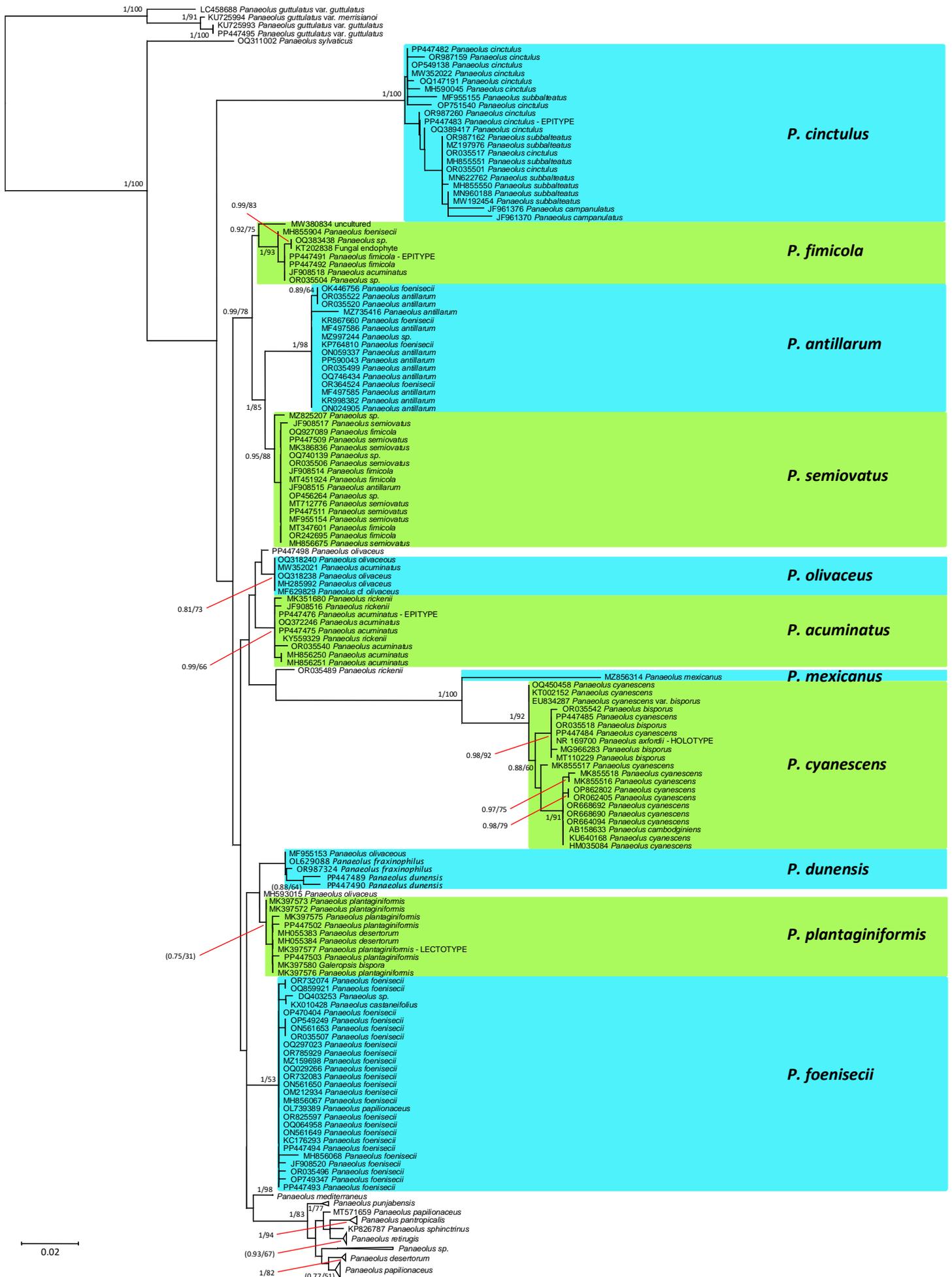
<b>species name in GenBank</b>	<b>current name</b>	<b>voucher/origin</b>	<b>ITS</b>
<i>P. papilionaceus</i>	<i>P. foeniseccii</i>	CIRM BRFM 715/?	OL739389
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	Mushroom Observer # 288725/Mexico	MH169580
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	FLAS-F-69055/USA	OP163246
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	FLAS-F-69481/USA Florida	OQ746433
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	DNA1940/USA Florida	KF830093
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	D15/China	KC414234
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	HFJAU0032/China	MN258670
<i>P. papilionaceus</i>	<i>P. pantropicalis</i>	MHHNU 31396/China	OP862800
<i>P. papilionaceus</i>	<i>P. punjabensis</i>	Shelly024/Namibia	UDB080025
<i>P. papilionaceus</i>	<i>P. punjabensis</i>	EGDA-Pan228/Egypt	MW915589
<i>P. papilionaceus</i>	<i>P. retirugis</i>	ECO-TA-HO 7877/Mexico	MF156263
<i>P. papilionaceus</i>	<i>P. retirugis</i>	iNAT:69899192/USA	MZ666372
<i>P. papilionaceus</i>	<i>P. retirugis</i>	NAMA 2017-161/USA	MH979305
<i>P. cf papilionaceus</i>	<i>P. retirugis</i>	MushroomObserver.org/312079/USA	MH101639
<i>P. papilionaceus</i> var. <i>capitatocystis</i>	<i>P. retirugis</i>	TUF118728/Estonia	UDB019537
<i>P. papilionaceus</i> var. <i>parvisporus</i> T	<i>P. papilionaceus</i>	Gerhardt 5.7.1988 (B)/Germany	PP447501
<i>P. papilionaceus</i> var. <i>parvisporus</i> P	<i>P. retirugis</i>	B700108103/Germany	<a href="#">PP590041</a>
<i>P. papilionaceus</i> var. <i>parvisporus</i> P	<i>P. retirugis</i>	B700108102/Germany	<a href="#">PP590042</a>
<i>P. papilionaceus</i> var. <i>parvisporus</i> P	<i>P. pantropicalis</i>	B700108104/Brazil	<a href="#">PP590040</a>
<i>P. papilionaceus</i> var. <i>retirugis</i>	<i>P. retirugis</i>	iNAT:21623086/USA	OM212937
<i>P. plantaginiformis</i>		LE 313092/Russia	MK397573
<i>P. plantaginiformis</i>		LE 2869/Russia	MK397572
<i>P. plantaginiformis</i> L		LE 2862/Russia	MK397577
<i>P. plantaginiformis</i>		LE 2870/Uzbekistan	MK397576
<i>P. plantaginiformis</i>		LE 2867/Uzbekistan	MK397575
<i>P. plantaginiformis</i>		TAAM120547/Uzbekistan	PP447502
<i>P. plantaginiformis</i>		TAAM120647/Uzbekistan	PP447503
<i>P. punjabensis</i> T		LAH 36793/Pakistan	NR189851
<i>P. punjabensis</i> P		LAH37417/Pakistan	OP681142
<i>P. punjabensis</i> P		BWN_45/Pakistan	MZ265143
<i>P. punjabensis</i>		ANGE1898/Dominican Republic	<a href="#">PP998475</a>
<i>P. retirugis</i>		CBS:272.47/France	MH856253
<i>P. retirugis</i>		CBS:273.47/France	MH856254
<i>P. retirugis</i>		CBS:274.47/France	MH856255
<i>P. retirugis</i>		CBS:324.34/?	MH855549
<i>P. retirugis</i>		7070/Italy	JF908521
<i>P. retirugis</i>		AMB 20077/Italy	PP447508
<i>P. retirugis</i>		AMB 20078/Italy	PP447505
<i>P. rickenii</i>	<i>P. acuminatus</i>	KA16-1041/Kyrgyzstan	MK351680
<i>P. rickenii</i>	<i>P. acuminatus</i>	TENN:054965/Argentina	KY559329
<i>P. rickenii</i>	<i>P. acuminatus</i>	749/Italy	JF908516
<i>P. rickenii</i>	<i>P. papilionaceus</i>	12446/Italy	JF908523

species name in GenBank	current name	voucher/origin	ITS
<i>P. rickenii</i>	<i>P. sp.</i>	4474/China	OR035489
<i>P. semiovatus</i>		AMB 20084/Italy	PP447509
<i>P. semiovatus</i>		AMB 20082/Italy	PP447511
<i>P. semiovatus</i>		4083/Italy	JF908517
<i>P. semiovatus</i>		GL-13/India	MK386836
<i>P. semiovatus</i>		GS2/China	OR035506
<i>P. semiovatus</i>		Mushroom Observer 377584/USA	MT712776
<i>P. semiovatus</i>		UBC F-23942/Canada	MF955154
<i>P. semiovatus</i>		CBS 388.50/France	MH856675
<i>P. semiovatus</i>	<i>P. retirugis</i>	CBS:276.39/?	MH856012
<i>P. sp.</i>	<i>P. fimicola</i>	BR5020211847626V/Belgium	OR035504
<i>P. sp.</i>	<i>P. foeniseccii</i>	705-2/China	DQ403253
<i>P. sp.</i>		iNat72986889/USA	OQ383438
<i>P. sp.</i>	<i>P. semiovatus</i>	LAH05071008/Pakistan	MZ825207
<i>P. sp.</i>	<i>P. semiovatus</i>	JLF9258/USA	OQ740139
<i>P. sp.</i>	<i>P. semiovatus</i>	ubco14/Canada	OP456264
<i>P. sp.</i>	<i>P. antillarum</i>	biocode08-94/French Polynesia	MZ997244
<i>P. sp.</i>	<i>P. punjabensis P**</i>	S1 (LAH36792)/Pakistan	KY636363
<i>P. sp.</i>	<i>P. papilionaceus</i>	PDD: 105318/New Zealand	MH380188
<i>P. sphinctrinus</i>	<i>P. papilionaceus</i>	CBS:582.79/?	HM035081
<i>P. sphinctrinus</i>	<i>P. papilionaceus</i>	CBS:582.79/France	MH873000
<i>P. sphinctrinus</i>	<i>P. papilionaceus</i>	TFB8627/Argentina	KY559331
<i>P. sphinctrinus</i>	<i>P. papilionaceus</i>	AFTOL-ID 1499/USA	DQ182503
<i>P. sphinctrinus</i>	<i>P. papilionaceus</i>	HMAS 290139/China	MK966651
<i>P. sphinctrinus</i>	<i>P. retirugis</i>	232/Italy	JF908513
<i>P. sphinctrinus</i>	<i>P. sp.</i>	UOC SIGWI S47/Sri Lanka	KP826787
<i>P. aff. sphinctrinus</i>	<i>P. sp.</i>	NY04449017/Colombia	<b>PP590035</b>
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	UBC F-23948/Canada	MF955155
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	OMDL K. Canan iNaturalist # 148029993/USA	OR987162
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	iNAT:16440988/USA	MZ197976
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	CBS 326.34/USA	MH855550
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	CBS 327.34/USA	MH855551
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	HFJAU-ND146/China	MN622762
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	4/China	MW192454
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	NX180911-04/China	MN960188
<i>P. subbalteatus</i>	<i>P. cinctulus</i>	/China	JF961370
<i>P. sylvaticus</i>		ANGE1393/Dominican Republic	OQ311002
undetermined	<i>P. sp.</i>	54A2/Colombia	MW380834
undetermined (fungal endophyte)	<i>P. sp.</i>	C111L/USA	KT202838

**Figure 1** (next page). Best scoring ITS rDNA phylogram of genus *Panaeolus* (with *P. guttulatus* as outgroup) obtained using RAxML. Nodes were annotated if they were supported by  $\geq 0.95$  Bayesian posterior probability (left) or  $\geq 70\%$  maximum likelihood bootstrap proportions (right). Nonsignificant support values are exceptionally represented inside parentheses. Sequences newly generated in this study are in bold.

New data and update in the *Panaeolus papilionaceus* complex, first record of *P. punjabensis* and new finds of *P. sylvaticus* in the Dominican Republic





## TAXONOMY

*Panaeolus punjabensis* M. Asif, Q. Firdous, A. Izhar, Niazi & Khalid (Fig. 2-7)

European Journal of Taxonomy 888: 85 (2023)

=? *Panaeolus alcis* (as '*alcidis*') sensu Kaur, Atri & Kaur (2014)

### Macroscopic characters

*Pileus* (primordia and young specimens not observed): 12 – 18 mm broad, paraboloid-convex; off-white to pale grey, at centre with a pale ochraceous-pinkish shade; cuticle dry, cracking and eroded with age, margin not striate; veil absent.

*Lamellae* adnate, spaced, approx 18 – 20, intermixed with 1-3 lamellulae, adnate, ventricose; at first olivaceous, then violaceous-bluish grey with blackish spots ("salt and pepper" appearance); edge pale.

*Stipe* 110 – 130 × 3 – 4.5 mm, base swollen up to 6.6 mm, cylindric, flexuous, straight to twisted; surface pale brownish with an olivaceous shade at apex, medium to darkish brown to reddish-violaceous brown elsewhere, distinctly covered in white pruina at apex and white fibrils elsewhere, base covered with white mycelial felt and floccules; bruising blue towards base; annulus absent.

*Context* not examined.



Fig. 2: *P. punjabensis*. ANGE188 [C. Angelini]



Fig. 3: *P. punjabensis*. ANGE188 [C. Angelini]



Fig. 4: *P. punjabensis*. ANGE188 [C. Angelini]

### Microscopic characters

*Basidiospores* (n=20) (14.0) 15.0 – 17.6 (19.2) × 9.0 – 10.2 (12.0) × 7.7 – 9.5 μm, Q 1.50 – 1.73 × 1.65 – 1.94; in front view broadly elliptic to broadly fusiform or subhexagonal, base obtuse to conical-obtuse, in side view elliptic-oblong; apex not or little protruding; dark brown to blackish brown in 5% KOH, distinctly thick-walled; germ pore distinct, 2.0 – 2.5 μm broad.

*Basidia* 24.0 – 34.0 × 10.0 – 13.7 μm, stoutly cylindrical to stoutly subululiform, rarely clavate, 4-spored; hyaline.

*Cheilocystidia* 35.0 – 50.0 μm long, base 5.0 – 11.5 μm broad, apex (3.0) 5.0 – 11.0 μm broad, lageniform, lageniform-clavate, clavate-to capitate-pedicellate, rarely cylindrical or tapering at apex, usually when swollen at apex then little ventricose at base; mostly hyaline and smooth, thin-walled, abundant. *Pleurocystidia* absent.

*Pileipellis* cellular composed of up to 30 μm broad elements.

*Pileocystidia* approx. 35 – 45 × 7 – 13 μm, lageniform, scattered.

*Clamp connections* found in pileus trama and at base of cheilocystidia.

**Collection examined and habitat:** Dominican Republic, P.to Plata, Cabarete, on grassy, heavily horse grazed ground of back dunes among shrubs of *Coccoloba uvifera*, 12 November 2023, legit C. Angelini (pers. herb. ANGE1898), GenBank: PP998475 - ITS.



Fig. 5: *P. punjabensis*. ANGE1898. Stipe base with bluish tints [C. Angelini]

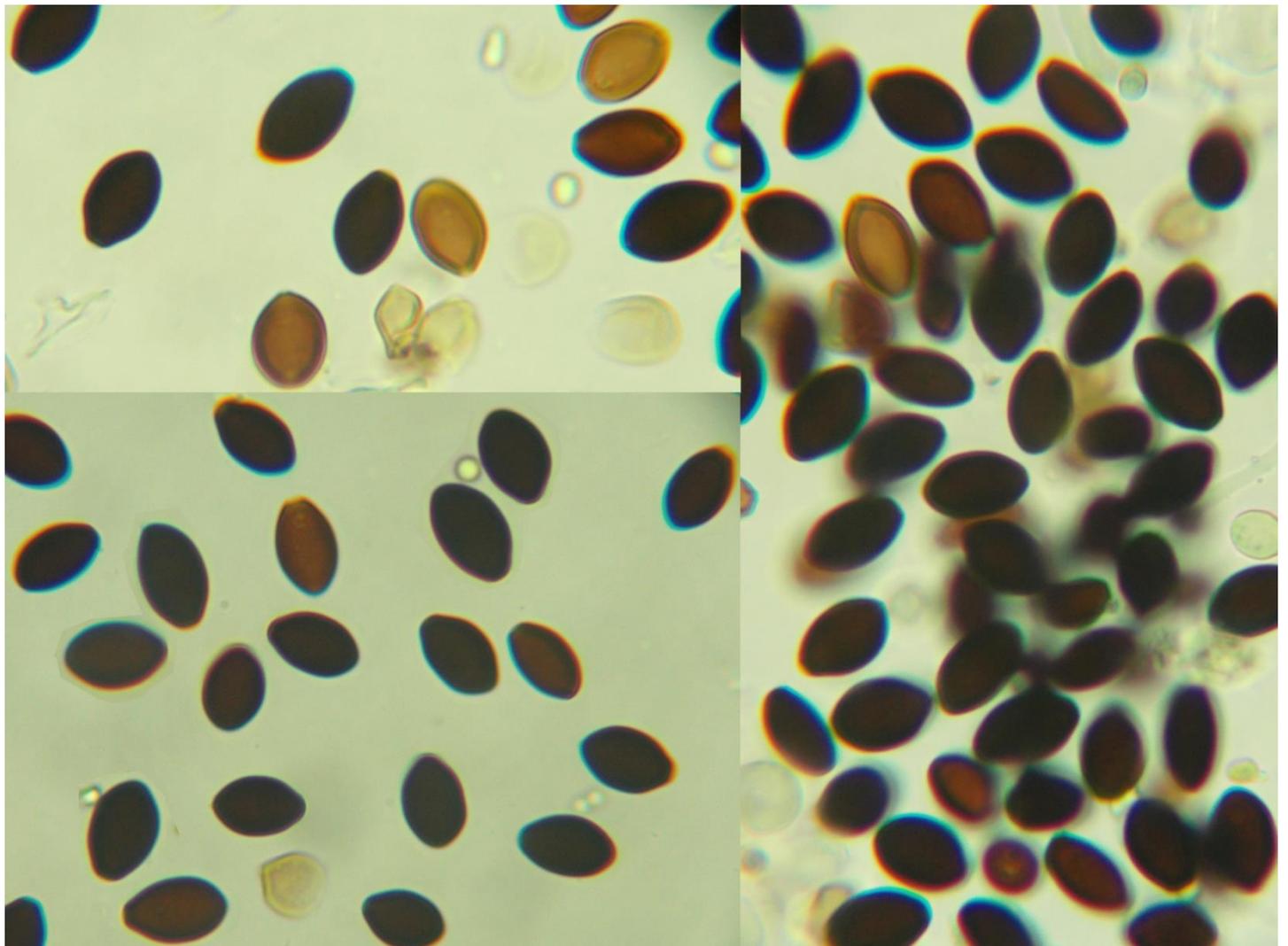


Fig 6: *P. punjabensis*. ANGE1898. Spores in 5% KOH [P. Voto]

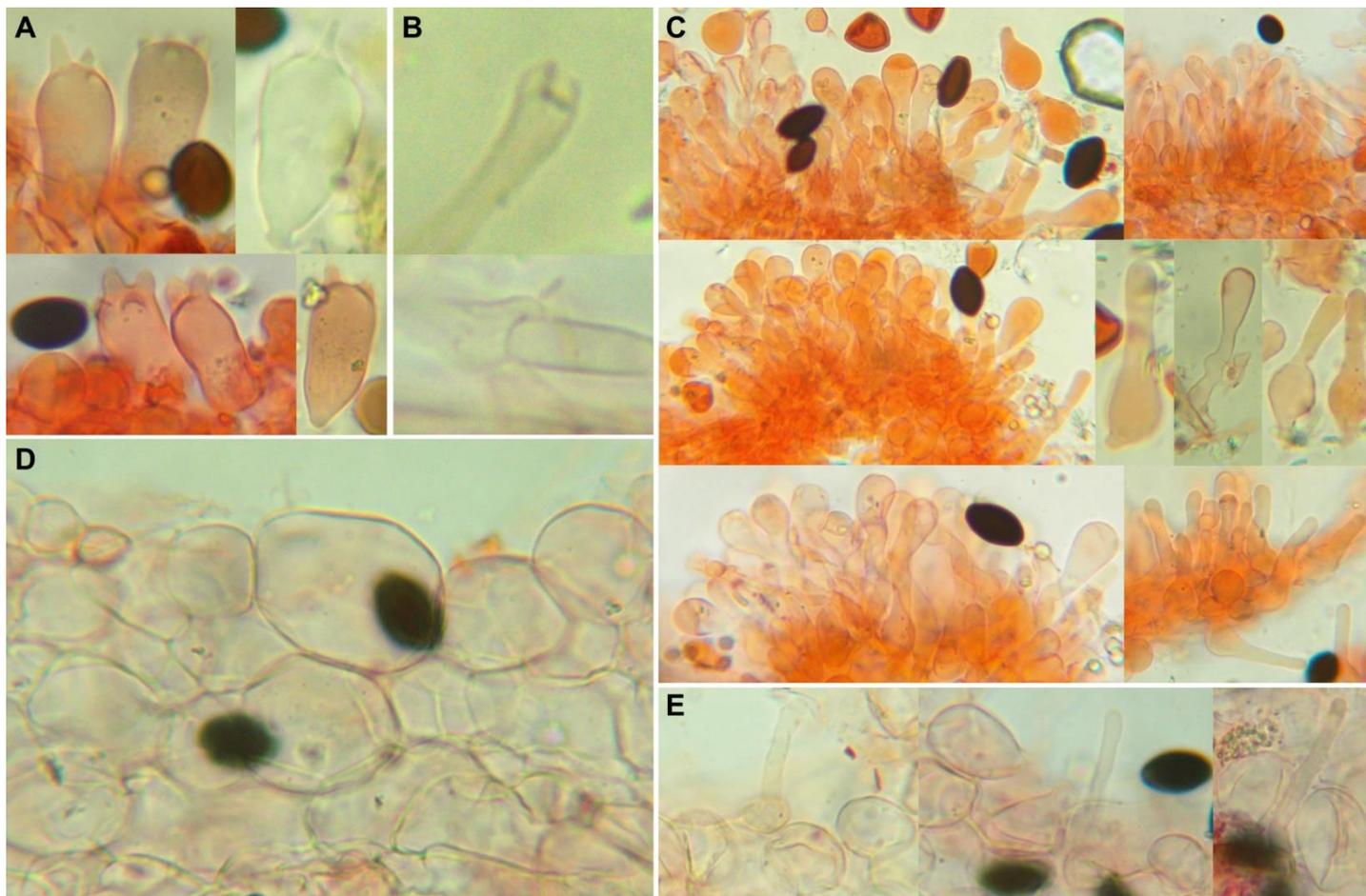


Fig 7: *P. punjabensis*. ANGE1898. A) Basidia, B) Clamps, C) Cheilocystidia, D) Pileipellis, E) Pileocystidia. All images in Congo red [P. Voto]

## Notes

Our description offers new relevant data integrating those reported in the protolog (Asif *et al.* 2023).

In particular we have noticed in our collection a not striate margin of pileus, a partial lentiform shape of the spores, narrower and 4-spored basidia, lageniform and scattered pileocystidia, presence of clamps.

*Panaeolus punjabensis* is characterized by absence of pleurocystidia (subgenus *Panaeolus*), agaricoid habit, absence of veil, whitish to pale grey pileus with ochraceous shades only at centre, and tending to crack and peel off, bluish tints on handling or bruising (we have not performed chemical tests attesting the presence of hallucinogenic substances), a mixture of lageniform to lageniform-clavate or claviform-capitate cheilocystidia, large elliptic-subfusiform, scarcely hexagonal-lentiform spores, habitat on nitrogenous soil with herbivorous dung present or directly on dung in tropical/subtropical climate.

Beyond the Indian region and the Caribbean, the taxon is also present, based on Genbank ITS sequences, in Egypt and in Namibia, evincing a broad diffusion.

Morphologically, the closest taxon seems to be *Panaeolus moellerianus* Singer, typified from the Macquarie Island in Oceania on humus among graminaceous plants, unsequenced. The pileus is defined as scrobiculate to reticulate-wrinkled (Singer 1960).

There are other unveiled, large-spored species in subgenus *Panaeolus*, they all differ by distinctly lentiform spores: *P. cinctulus* (Bolton) Sacc. (with also a partially eccentric germ pore), *P. acuminatus* (P. Kumm.) Quél., *P. paludosus* Cleland and *P. goossensiae* Beeli.

Finally, we report an unsequenced collection identified as *P. alcis* (as '*alcidis*') by Kaur, Atri & Kaur M (2014), from India. This material features all characteristics we found in our collection. The authors report having noticed a bluing also on the pileus and in the context. We are convinced this represents a misidentification of *P. punjabensis*.

## NEW DATA IN THE *PANAEOLUS PAPILIONACEUS* COMPLEX

An ITS sequence of the holotype of *Panaeolus papilionaceus* var. *parvisporus* has very recently become available (it was repeatedly and unsuccessfully tried by Voto & Angelini 2024). Since the phylogram demonstrates that it nests in the clade *Panaeolus papilionaceus* s.str. (see Fig. 1), the systematics of the *Panaeolus papilionaceus* complex as proposed in Voto & Angelini (2024) needs to be reassessed.

Voto & Angelini (2024) had phylogenetically demonstrated the presence in Europe of two taxa in the *P. papilionaceus* complex (ignoring the galeroid taxa).

Basing on the ITS sequences obtained from two German paratypes of *P. papilionaceus* var. *parvisporus*, they assumed that these two taxa could be separated on the base of the spore length: a large-spored (more than 15 µm long on average) taxon identified as *P. papilionaceus* (= *P. campanulatus* = *P. retirugis* = *P. sphinctrinus*) and a small-spored (at most 15 µm long on average) taxon they named *P. parvisporus* comb. nov. (the choice of the names followed the systematics proposed by Gerhardt 1996, 2012).

With the new molecular evidence of a small-spored collection occurring in the clade of *P. papilionaceus* s. str., the separation based on the spore length would become incorrect. The only morphological difference between these two taxa would therefore be based on the tendency towards a smooth and more greyish pileus in *P. papilionaceus* compared to a corrugated and more brownish pileus in *P. retirugis*.

However, we have examined the three following considerations.

a) Over time, several authors (e.g. Hora 1957; Ota 1970; Watling & Gregory 1987; Bon & Courtecuisse 2003) having treated the *Panaeolus papilionaceus* complex never reported small-spored collections combined with a smooth, non-corrugated pileus;

b) In all descriptions of the above mentioned authors the pileus breadth of *P. papilionaceus* falls in the range 20-45 mm. Therefore, a pileus 8-15 mm broad, as described by Gerhardt (1996) for *P. papilionaceus* var. *parvisporus*, is unusually very small;

c) In Bulliard's (1782, table 58) iconography of *Agaricus papilionaceus* (see reproduction in Voto & Angelini 2024 figure 19) mature pilei are drawn conical-convex to convex, and in Fries's (1821) sanction they are defined convex. On the contrary, Gerhardt (1996) defines the pileus of *P. papilionaceus* var. *parvisporus* as thimble-shaped to campanulate.

Ultimately, basing on these observations, we propose to treat the type collection of *P. papilionaceus* var. *parvisporus* as consisting of still immature specimens and therefore with spores not yet of the definitive size.

On this assumption, we confirm Voto & Angelini's (2024) proposal of using the spore length for separating the two taxa, we add to it the pileus characteristics and we renounce the use of the epithet '*parvisporus*' instead of '*retirugis*'.

The two European non-galeroid taxa of the *P. papilionaceus* complex are therefore:

- *P. papilionaceus* (= *P. campanulatus* = *P. sphinctrinus*) with spores on average more than 15 µm long, and pileus smooth and mostly in the greyish tones;

- *P. retirugis* (= *P. papilionaceus* var. *parvisporus* = *P. parvisporus*) with spores on average at most 15 µm long, and pileus corrugated and mostly in the brownish tones.

Pileus colour and corrugation of course may depend on environmental conditioning and the freshness of the basidiomata.

The tree used in Voto & Angelini's (2024) is here reproduced expanded with the sequence of the typus of *P. papilionaceus* var. *parvisporus* and some others and with the names of the species updated.

*Panaeolus sylvaticus* Silva-Filho & Cortez (Fig. 8-10)

Edinburgh Journal of Botany 76(2): 303 (2019)

**Collections examined and habitat.** Dominican Republic, P.to Plata, Cabarete, Sea Horse Ranch, gregarious on woody debris between piles of decaying plant material (landfill of material from pruning) in an almond forest near the beach, 4 February 2024, legit C. Angelini, pers. herb. ANGE1992; ibid, pers. herb. ANGE1993.



Fig. 8: *P. sylvaticus*. ANGE1992 [C. Angelini]



Fig. 9: *P. sylvaticus*. ANGE1993 [C. Angelini]

## Notes

Our two new collections feature morphological data fully correspondent to those already reported in Angelini & Voto (2023). We can only add that we found occasional larger spores attaining to 11.1x8.1x5.8  $\mu\text{m}$  and somewhat more evident greenish-yellowish mucus or content of cheilocystidia (Fig. 10). The photographs here published show the colours in discoloring pilei.

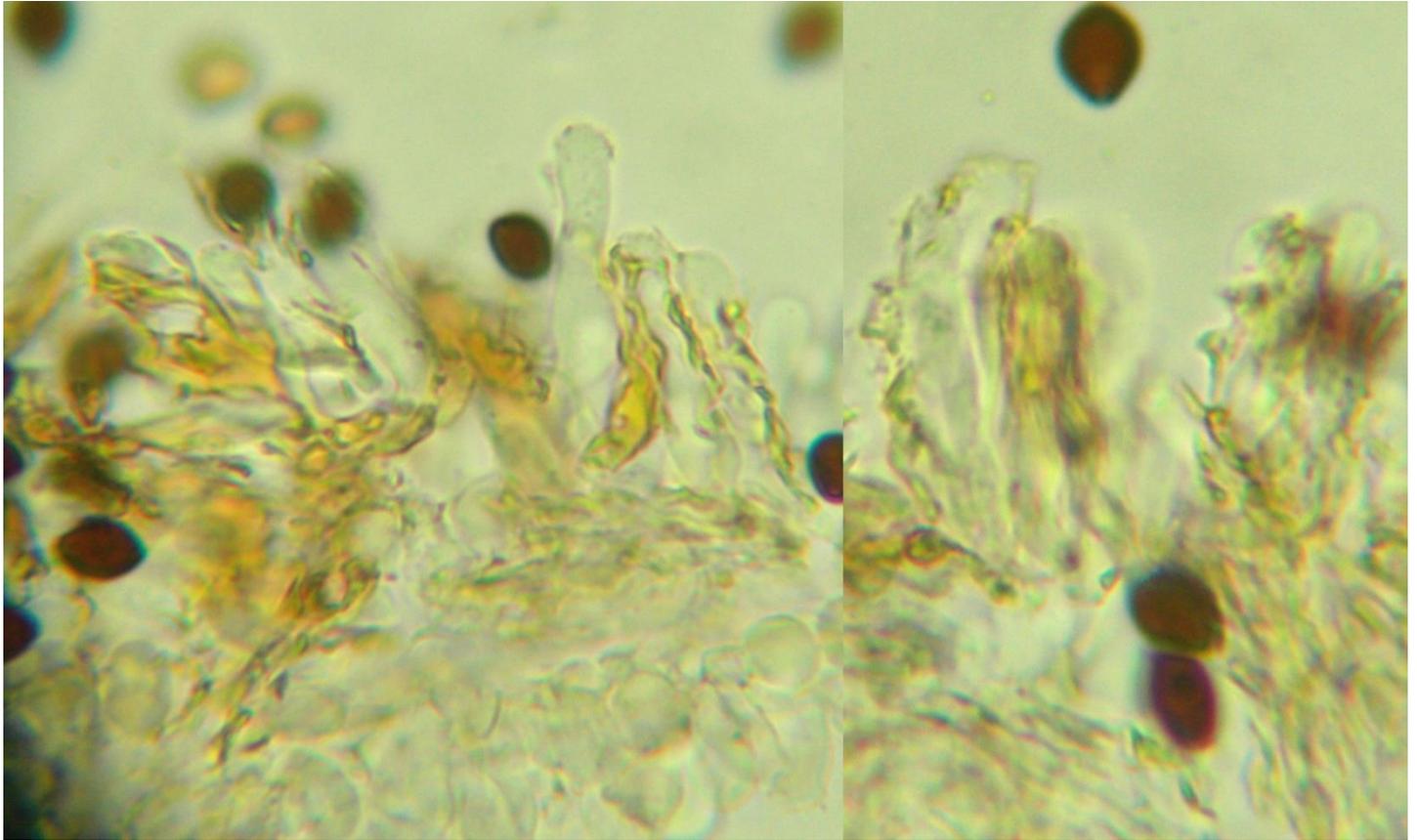


Fig. 10: *P. sylvaticus*. ANGE1992. Cheilocystidia in 5% KOH

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